

Use of primary conjugated antibodies to model the heterogeneity of GBM's

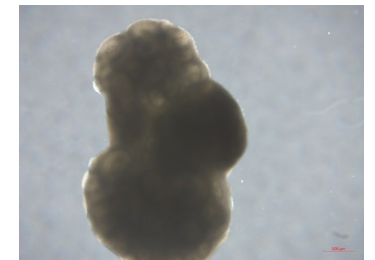
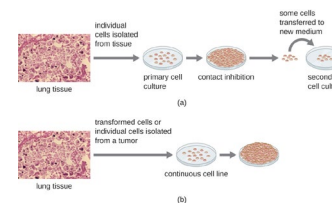
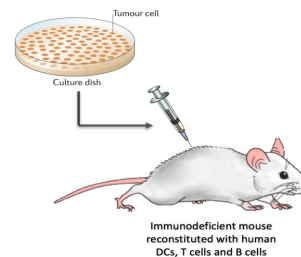
Ryan Gensler
NSSSP 2019



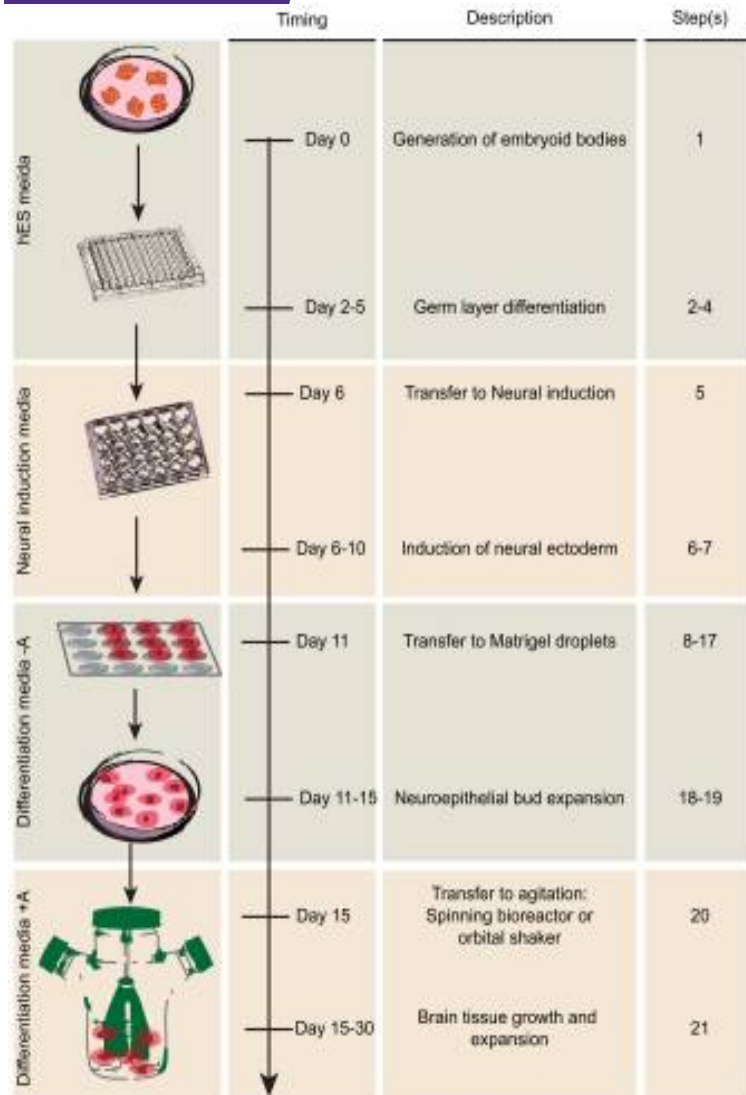


3 Models for GBM

Model Type	Xenografts	Cell Lines	Organoids
Pros	<ul style="list-style-type: none">• Heterogeneity• 3D morphology<ul style="list-style-type: none">• In vivo	<ul style="list-style-type: none">• Cheap• Grow quickly	<ul style="list-style-type: none">• Heterogeneity<ul style="list-style-type: none">• Cheap• 3D morphology• Act as patient avatars
Cons	<ul style="list-style-type: none">• Expensive• Slow tumor growth	<ul style="list-style-type: none">• Lack heterogeneity<ul style="list-style-type: none">• In vitro	<ul style="list-style-type: none">• In vitro• Not yet verified



What/How are organoids grown?



Lancaster and Knoblich, Nat Protoc 2014

- **Organoid Morphology:**
 - > 3D
 - > Populated by neuronal, glial, and NSCs
 - > Organized tissue with temporal patterning
- **Start from ESCs and correct factors cause neural differentiation**
- **At day 30, inject tumor cells into the organoid**
- **Day 60-90: Organoid sectioned, stained, imaged**



Aim of Project

- Create primary conjugated antibodies
- Use these 1° conjugated antibodies to avoid the lengthy TSA-IHC protocol
- Co-stain tissue
- Use these co-stains to understand the heterogeneity of the GBMs
 - > I.e. what proteins GBMs are expressing for a specific patient tumor
- Big Picture: Prove that GBM injected tumors cells are initiating GBM growth in organoid models
- Compare organoid models to heterogeneity of the patient tumor



Methods

Unconjugated 1° antibodies



Conjugation dyes
15 minutes
Mistakes

Conjugated 1° antibodies
(Sox 2, Ki 67, Vimentin,
Ptpz 1, Hop X, olig 1)

Test for optimal antibody
to solvent concentration

Find optimal
concentrations

Select random slide for
testing

Samples frozen in OCT



Cold cryostat
18 uM
Box of slides

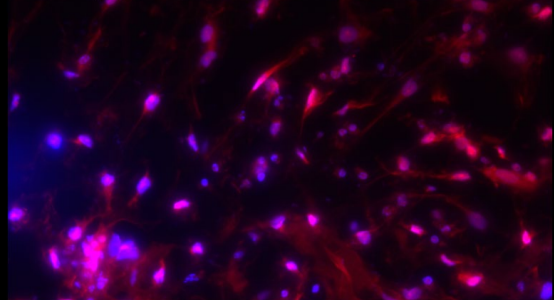
30-50 slides of sectioned
tissue stored at -80°C

Ptpz1

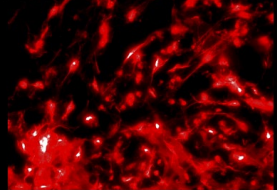
HOPX

Vimentin

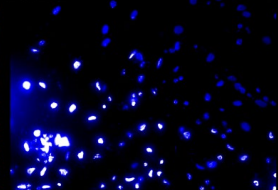
C19-27 w/o pdgf



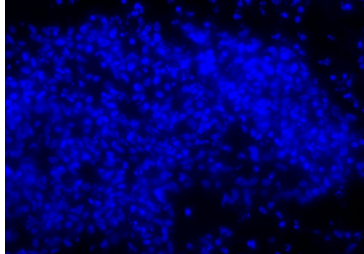
PTPRZ1



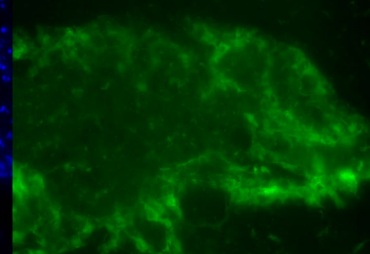
DAPI



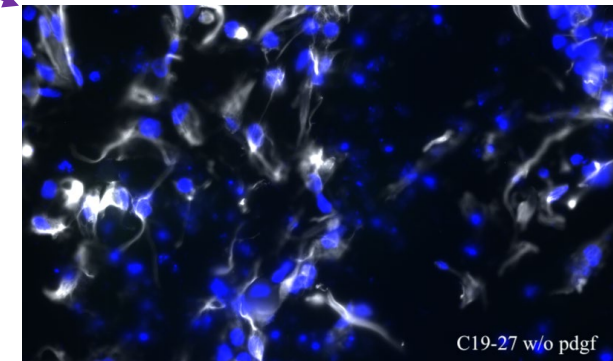
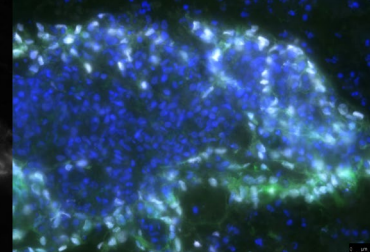
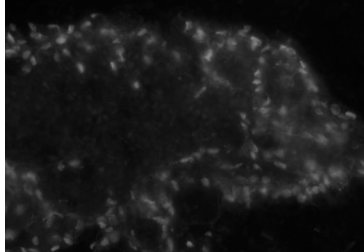
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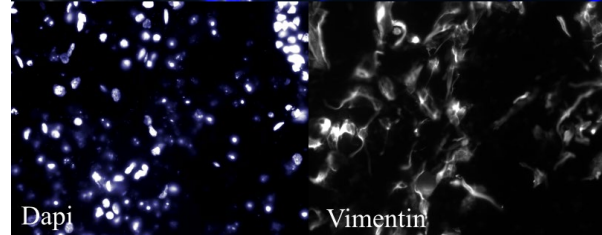
GFP



HOPX



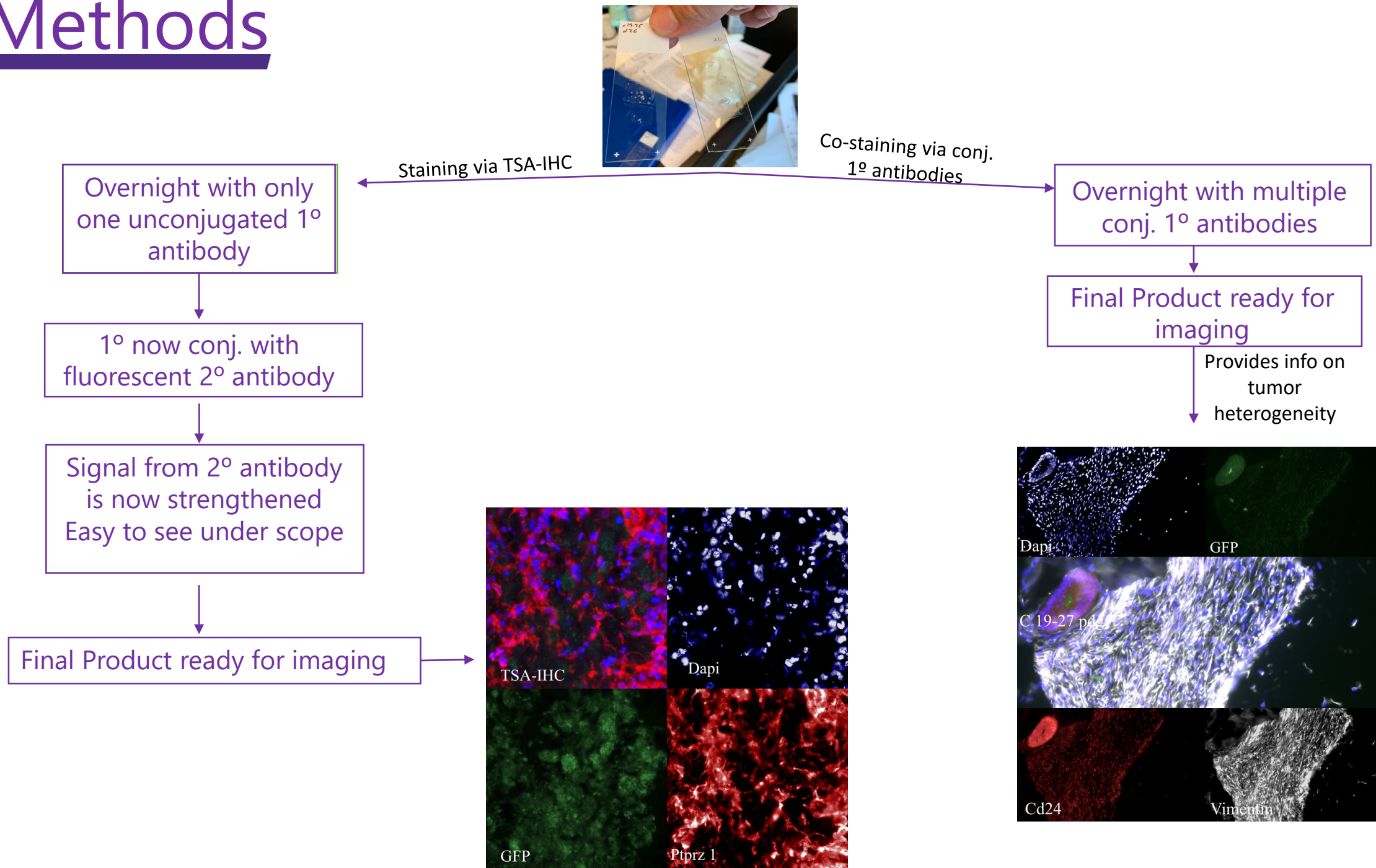
C19-27 w/o pdgf



Dapi

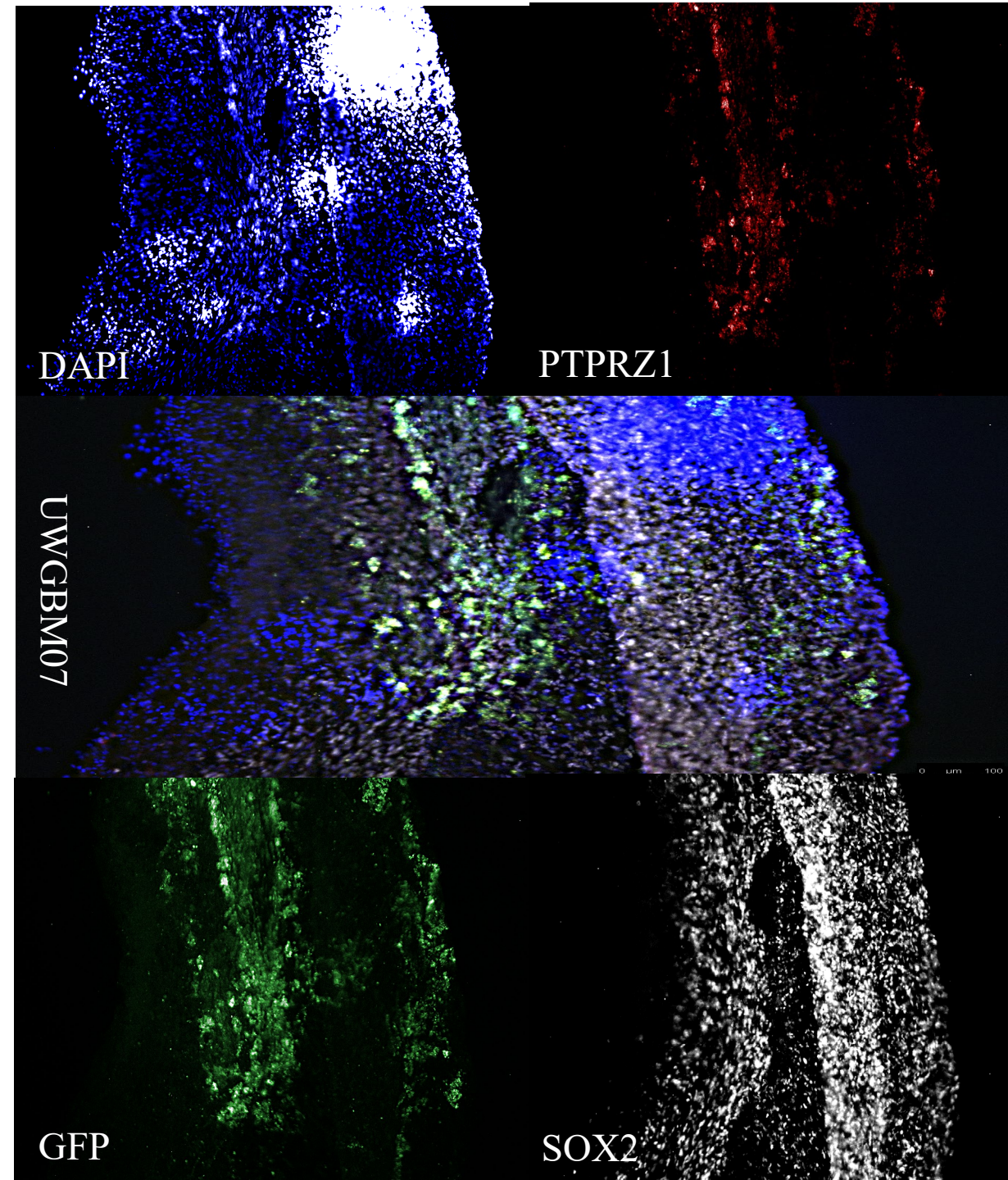
Vimentin

Methods



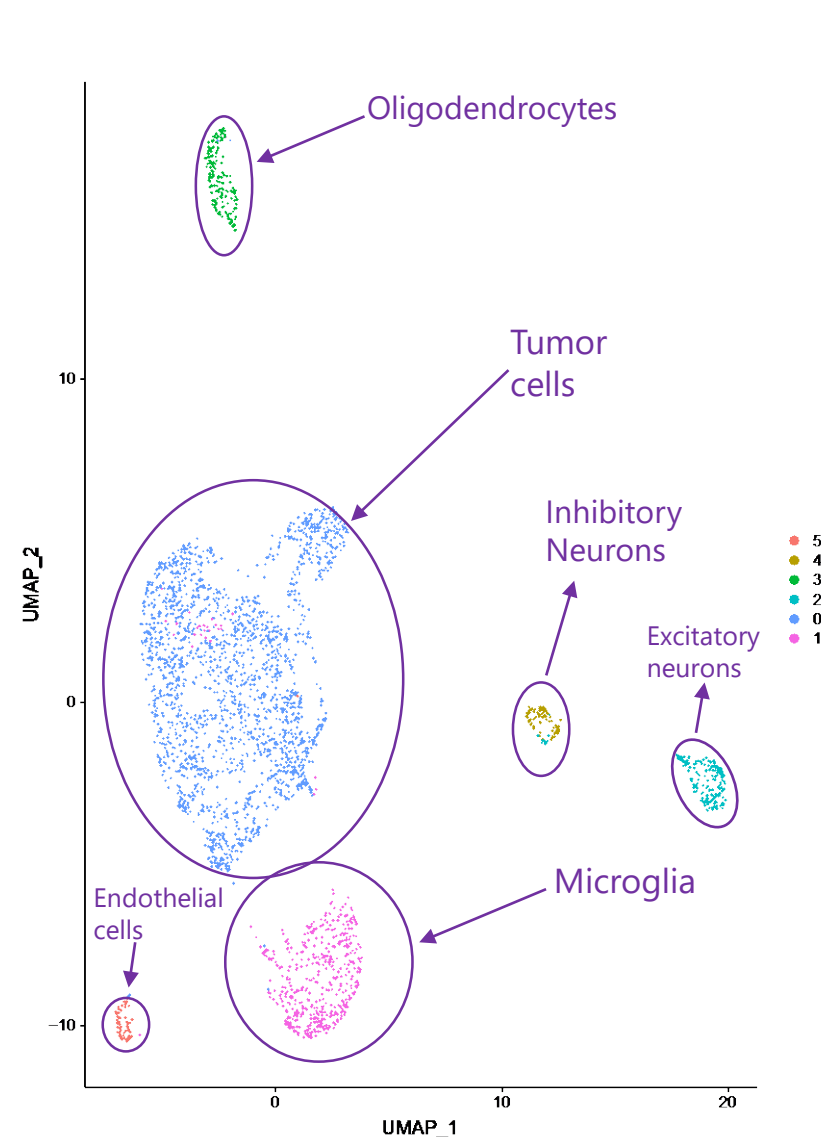
Proving Tumor Cells are in Organoids

- Validated staining protocol
- Stained for all tumor cells (SOX2)
- Would expect organoids to be sox 2 and ptpnz1 , but not GFP
- Next step: Compare heterogeneity patient tumor to organoid via scRNA-seq

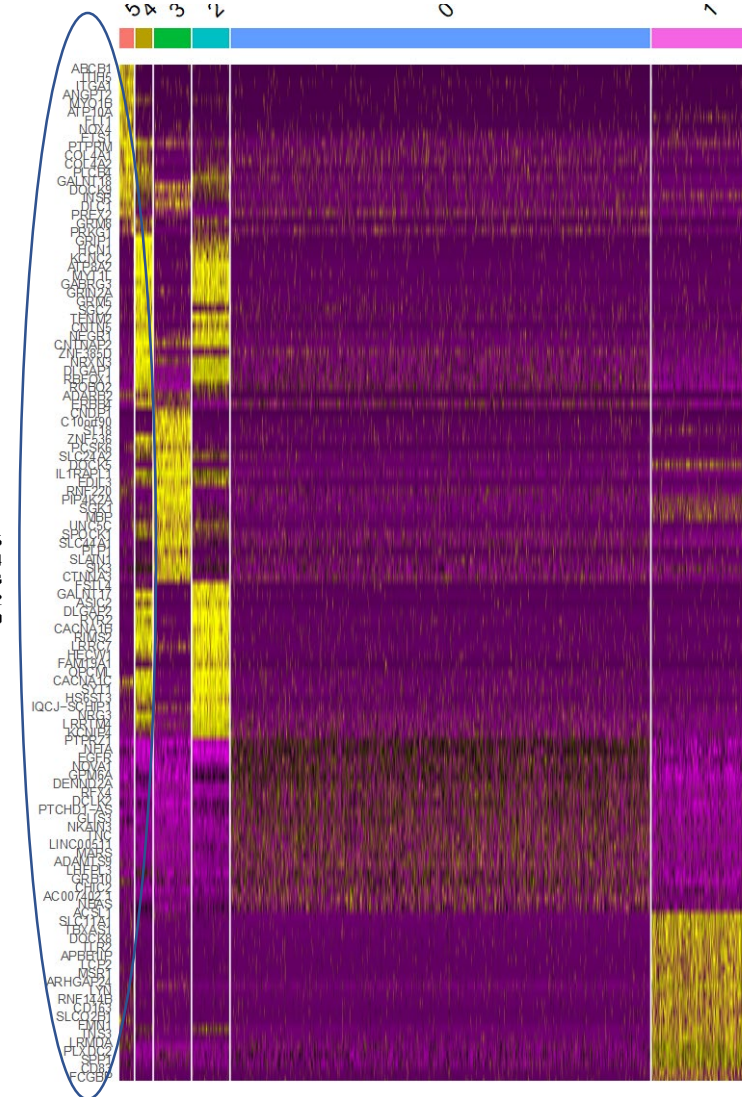




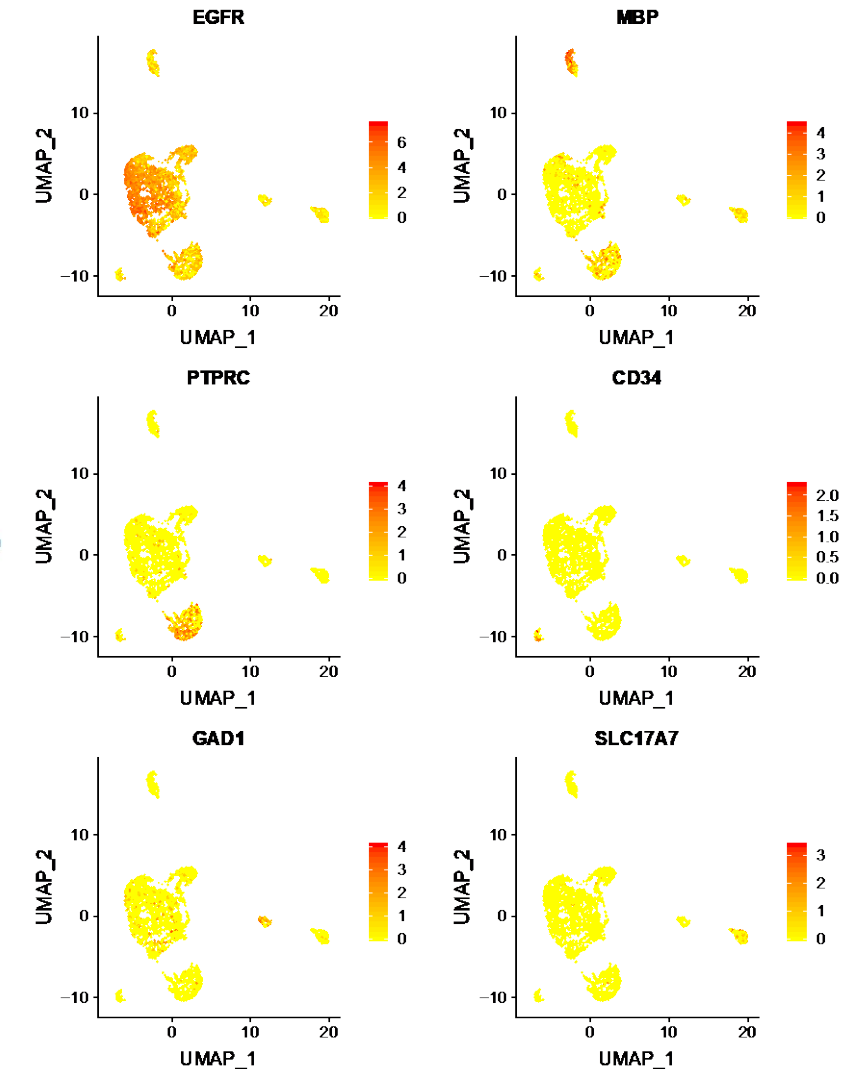
Comparing sc-RNAseq to Stains



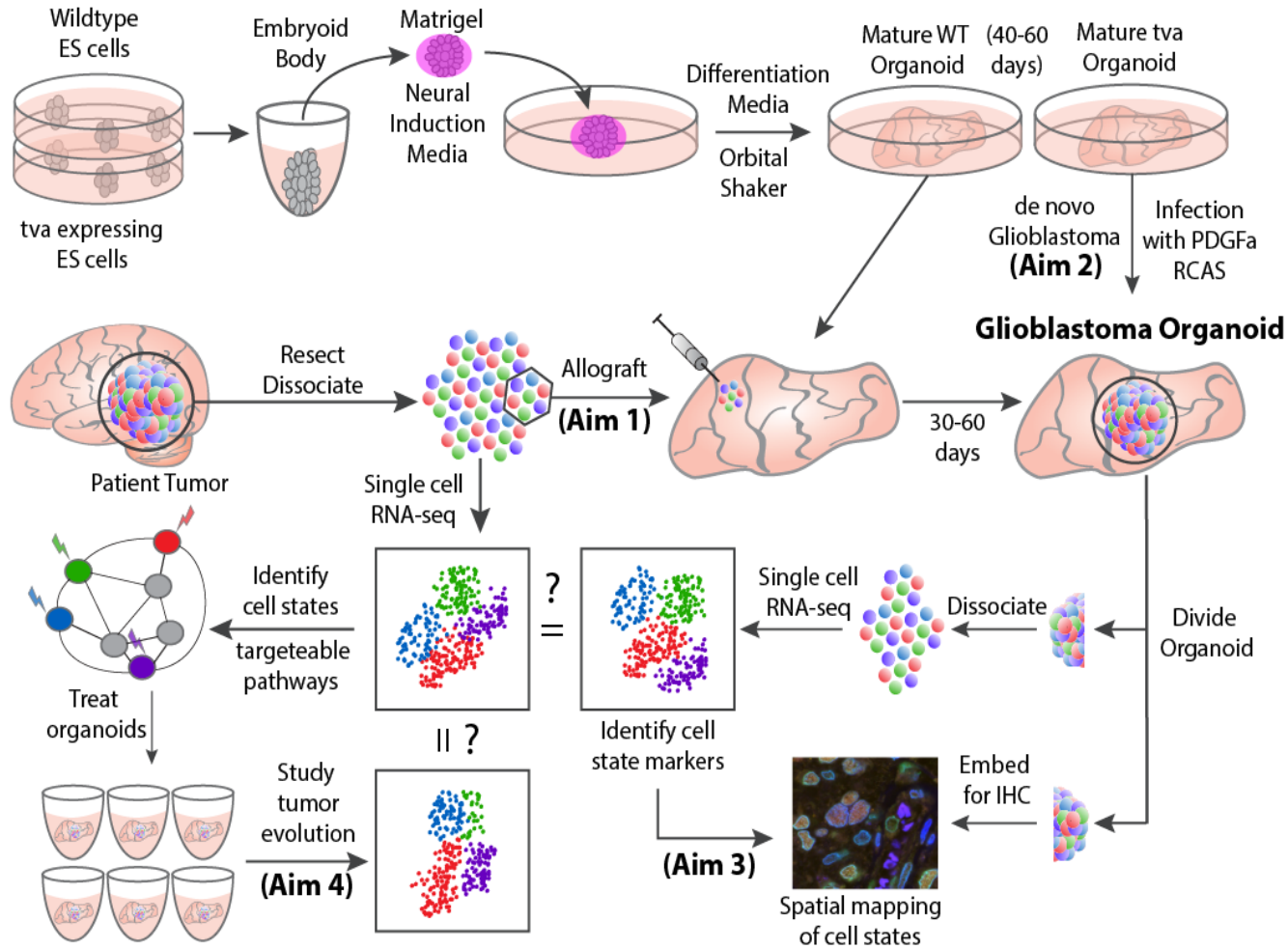
Umap graph shows clustering of similar cells.



Heat map showing groups of cells that express similar genes



Future Ventures



Goals:

1. Improve allograft injection method of organoid with human GBM cells
2. Use FISH to ultimately identify human tissue in organoids
3. Use scRNA-Seq to sequence organoids to prove model viability
4. Continue to use staining method to visualize heterogeneity of GBMs embedded in organoids

Figure by Anoop Patel

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